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lung (NCI-H460), and CNS (SF-268) cancer cell lines.

The complex molecular architecture and rich functionalization of marine natural products make them utterly attractive

targets for total synthesis.¹ In particular, marine diterpenes isolated from the large variety of gorgonian species (sea whips, sea feathers, sea plumes, and sea fans) found within the Caribbean Sea region have for many years captured the attention of synthetic and biosynthetic chemists alike.² As part of our continuing chemical investigation of Caribbean gorgonian octocorals, we report here the isolation, structure

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Isolation and Structure of Providencin: A Highly Oxygenated Diterpene Possessing a Unique Bicyclo[12.2.0]hexadecane Ring System from the Sea Plume Pseudopterogorgia kallos

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determination, and biological activity of a novel, highly oxygenated polycyclic diterpene, providencin (1).

The conspicuous sea plume Pseudopterogorgia kallos (Bielschowsky, 1918) was collected near Providencia (Old





Providencin (1) is a naturally occurring cytotoxin isolated from the Caribbean gorgonian octocoral Pseudopterogorgia kallos. Its highly oxygenated hexacyclic structure is based on a previously undescribed bicyclo[12.2.0]hexadecane ring system and was established through spectroscopic analysis and X-ray crystallographic analysis. Providencin (1) was shown to exhibit modest anticancer activity against human breast (MCF7),

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ABSTRACT

Fable 1. ¹ H NMR (300 MHz), ¹³ C NMR (75 MHz), ¹ H $^{-1}$ H COSY, NOESY, and HMBC Spectral Data for Providencin (1) ^{<i>a</i>}					
position	$\delta_{ m H}$, mult (J in hertz)	$\delta_{ m C}$ (mult)	¹ H ⁻¹ H COSY	NOESY	HMBC ^b
1	4.68, m	43.1 (CH)	H2, H14 α , H16 $\alpha\beta$	$H14\beta$	H2, H13, H14αβ, H16αβ, H17
2	3.47, dd (10.0, 6.1)	42.7 (CH)	H1, H17	H13, H14α, H17	Η14α, Η17
3		157.9 (C)			H2, H5
4		114.3 (C)			H2, H5
5	6.53, d (1.2)	107.8 (CH)	H7		H7
6		149.0 (C)			H5, H7
7	4.03, br s	55.1 (CH)	H5	$H9\beta$	H9α, H ₃ -19
8		56.0 (C)			H7, H9α $β$, H10, H ₃ -19
9α	2.45, dd (15.3, 2.8)	39.4 (CH ₂)	H9β, H10	H10, H11, H ₃ -19	H7, H10, H ₃ -19
9β	2.05, dd (15.3, 4.0)		Η9α, Η10	H7, H10	
10	4.76, dd (4.0, 2.8)	75.7 (CH)	Η9αβ	H9αβ, H11	H9β, H11
11	3.99, s	60.6 (CH) ^c		H9α, H10, H ₃ -19	H9β, H10, H13
12		58.1 (C) ^c			H10, H11, H14α
13	5.45, br d (10.9)	65.2 (CH) ^c	$H14\beta$	H2, H14α, H ₃ -19	Η14α
14α	1.73, m	34.7 (CH ₂)	H1, H14 β	H2, H13	H2, H13
14β	3.02, m		Η13, Η14α	H1	
15		151.0 (C)			H14α, H16αβ, H17
16α	5.21, br d (3.1)	108.6 (CH ₂)	H1, H16 β	H16 β	
16β	5.05, br d (2.2)		Η1, Η16α	Η16α	
17	5.06, br d (6.1)	72.8 (CH)	H2	H2	H2, H16 $\alpha\beta$
18		164.7 (C)			H ₃ -21
19	1.18, s	20.1 (CH ₃)		H9α, H11, H13	H9 β
20		167.9 (C)			H11
21	3.83, s	51.9 (CH ₃)			
22		170.0 (C)			H13, H ₃ -23
23	2.06, s	20.8 (CH ₃)			

^{*a*} Spectra were recorded in CDCl₃ at 25 °C. Chemical shift values are in parts per million relative to TMS. ¹³C NMR multiplicities were obtained from a DEPT-135 experiment. ^{*b*} Protons correlated to carbon resonances in ¹³C column. Parameters were optimized for ^{2,3} $J_{CH} = 6$ and 8 Hz. ^{*c*} Broad low-intensity resonance line.

Providence) Island located in the Southwestern Caribbean Sea, among the oceanic islands, atolls, and banks of the San Andrés and Providencia Archipelago, off the Nicaraguan shelf.³ The sun-dried animal specimens (1.07 kg) were frozen, cut in small pieces, and homogenized exhaustively using a mixture of CH₂Cl₂-MeOH (1:1). After in vacuo concentration, the dried extract (166 g) was subjected to our standard partitioning procedure, resulting in a hexane, CHCl₃, and EtOAc fraction.⁴ The CHCl₃-soluble material (39.3 g) was purified by repeated silica gel flash chromatography and size-exclusion chromatography on a Bio-Beads SX-3 column to yield 20 mg (0.012% dry wt) of pure providencin (1). The structure of the new cytotoxin was defined by ¹H NMR, ¹³C NMR, DEPT, ¹H-¹H COSY, HMQC, HMBC, and twodimensional NOESY experiments. A single-crystal X-ray structure analysis was subsequently carried out in order to confirm the proposed molecular structure of 1.

Providencin (1), $[\alpha]^{20}{}_{\rm D}$ +7.9° (*c* 1.2, CHCl₃), was isolated as a colorless amorphous solid whose molecular formula of C₂₃H₂₄O₁₀ was established on the basis of HREIMS analysis ([M]^{+•}; *m/z* 460.1366, +0.0004 mmu),⁵ ¹³C NMR, and DEPT

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spectra. Thus, 12 degrees of unsaturation were determined for 1. The IR spectrum of 1 indicated the presence of hydroxyl (ν_{max} 3461 cm⁻¹), ester (ν_{max} 1783, 1743, 1718 cm⁻¹), olefin (ν_{max} 3129, 3078, 1648, 1620, 1577 cm⁻¹), and epoxide (ν_{max} 1235 cm⁻¹) groups, and the UV spectrum (MeOH) showed maxima at λ_{max} 203 (ϵ 11500) and 251 (ϵ 4900) nm. NMR experiments were performed in CDCl₃ to enhance peak separation; however, partial peak broadening in the ¹H NMR as well as peak splitting in the ¹³C NMR spectra was observed, suggesting the presence of at least two distinct conformations in solution (Table 1). The ¹³C NMR spectrum displayed signals for 23 carbons. DEPT analysis indicated that 1 contained three methyls, three methylenes, eight methines, and nine quaternary carbons. The presence of nine sp²-hybridized carbon atoms in the molecule, as deduced from the ¹³C and DEPT NMR spectra, corresponding to three carbon-carbon and three carbon-oxygen double bonds as the only multiple bonds, indicated compound 1 to be hexacyclic. An HMQC experiment established all singlebond ¹H-¹³C connectivities.

The ¹H NMR spectrum (CDCl₃, 300 MHz), while being generally consistent with the presumption that **1** was a diterpene, showed only three methyl signals, all of which were sharp singlets displaced at $\delta_{\rm H}$ 3.83, 2.06, and 1.18, designated as H₃-21 (a carbomethoxyl), H₃-23 (an acyloxy methyl), and H₃-19. HMQC correlation of $\delta_{\rm H}$ 5.21 (br d, J = 3.1 Hz, H₂-16 α) and 5.05 (br d, J = 2.2 Hz, H₂-16 β)

⁽³⁾ For the only in-depth chemical investigation of *P. kallos* prior to this work, see: Look, S. A.; Burch, M. T.; Fenical, W.; Qi-tai, Z.; Clardy, J. *J. Org. Chem.* **1985**, *50*, 5741–5746.

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⁽⁵⁾ EIMS *m*/*z* 460 [M]⁺ (29), 442 (100), 400 (75), 382 (36), 367 (49), 218 (69), 167 (98), 136 (90), 90 (31).

with δ_{C} 108.6 (t, C16) and HMBC correlation (Table 1) of these protons with $\delta_{\rm C}$ 151.0 (s, C15) indicated that 1 contained a strained exocyclic methylene.⁶ Eight oxygenated carbon atoms were observed in the ¹³C NMR spectrum (CDCl₃, 75 MHz) of 1 in the range 51.9-75.7 ppm, which were assigned to three ester functionalities (an acetate, a γ -lactone, and a carbomethoxyl group), two epoxides, and a hydroxyl linked to a secondary carbon. The acetate was readily identified by the methyl resonance observed in the ¹³C and ¹H NMR spectra at $\delta_{\rm C}$ 20.8 (q, C23) and $\delta_{\rm H}$ 2.06 (s, 3H, H₃-23), respectively, while the other two ester groups were determined to be a conjugated carbomethoxyl and a nonconjugated γ -lactone, respectively, on the basis of NMR data at $\delta_{\rm C}$ 164.7 (s, C18), 51.9 (q, C21); $\delta_{\rm H}$ 3.83 (s, 3H, H₃-21) and $\delta_{\rm C}$ 167.9 (s, C20), 75.7 (d, C10); $\delta_{\rm H}$ 4.76 (dd, J = 4.0, 2.8 Hz, H-10). The ¹³C NMR singlets at $\delta_{\rm C}$ 157.9 (C3), 149.0 (C6), and 114.3 (C4) and a doublet at $\delta_{\rm C}$ 107.8 (C5) that was correlated in the HMQC experiment with the ¹H NMR signal at $\delta_{\rm H}$ 6.53 (d, J = 1.2 Hz, H-5), together with the carbomethoxyl signal at $\delta_{\rm H}$ 3.83 (s, 3H, H₃-21) in the ¹H NMR spectrum and at $\delta_{\rm C}$ 164.7 (s, C18) in the ¹³C NMR spectrum, were assigned to a α, α' -disubstituted β -carbomethoxyfuran constellation.⁷ HMQC correlation of $\delta_{\rm H}$ 4.76 (dd, J = 4.0, 2.8 Hz, H-10) with $\delta_{\rm C}$ 75.7 (d, C10) and of $\delta_{\rm H}$ 3.99 (s, H-11) with $\delta_{\rm C}$ 60.6 (d, C11), along with HMBC correlations of H-10 with $\delta_{\rm C}$ 60.6 (d, C11) and 58.1 (s, C12) and of H-11 with $\delta_{\rm C}$ 75.7 (d, C10), 58.1 (s, C12), and 167.9 (s, C20), indicated the presence of a α,β -epoxy γ -lactone moiety.8 The presence of a trisubstituted epoxide was evident from the ¹³C NMR singlet at $\delta_{\rm C}$ 56.0 (C8) and a doublet at $\delta_{\rm C}$ 55.1 (C7) that was correlated in the HMQC experiment with the ¹H NMR signal at $\delta_{\rm H}$ 4.03 (br s, H-7). The latter proton was correlated in the HMBC experiment with $\delta_{\rm C}$ 149.0 (s, C6) and 107.8 (d, C5). The methyl at $\delta_{\rm H}$ 1.18 (s, H₃-19) showed HMBC correlations with $\delta_{\rm C}$ 56.0 (s, C8), 55.1 (d, C7), and 39.4 (t, C9), indicating 1 contained a methylbearing trisubstituted epoxide flanked by a methylene carbon and a quaternary olefinic carbon. Thus, it quickly became apparent that 1 contained many of the same functionalities present in the bipinnatins, a series of highly oxygenated cembrane-based diterpenes isolated from the gorgonian Pseudopterogorgia bipinnata.9 On the other hand, two conspicuous methine carbon atoms at $\delta_{\rm C}$ 43.1 (d, C1) and 42.7 (d, C2), which were correlated by HMQC with proton signals directly correlated by ${}^{1}\text{H}{-}{}^{1}\text{H}$ COSY at δ_{H} 4.68 (m, H-1) and 3.47 (dd, J = 10.0, 6.1 Hz, H-2), respectively, were located in a cyclobutane ring along with the exocyclic double bond and secondary hydroxyl group. Additional ¹H-¹H COSY correlations observed between H-2 and the signal at $\delta_{\rm H}$ 5.06 (br d, J = 6.1 Hz, H-17), as well as long-range couplings between the terminal methylene protons H2-16 and H-1, established almost all of the connectivity about the cyclobutane ring. Further analysis of the HMBC experiment

(Table 1) provided additional connectivities between all six of the rings within **1** to form a bicyclo[12.2.0]hexadecane system (Figure 1). Some key HMBC correlations establishing



Figure 1. Trans-fused bicyclo[12.2.0]hexadecane ring system demarcates the novel providenciane carbon skeleton in providencin (1).

this ring system were observed between H-2 to C1, C3, C4, and C17; H-7 to C5, C6, C8, and C9; H-10 to C8, C9, C11, and C12; and H-13 to C1, C11, and C14. Connections from the cyclobutane system to the macrocycle were limited to HMBC correlations from H-2 to C1 and C17 and from H-16 α and H-16 β to C1, C15, and C17.

The relative stereochemistry of providencin (1) containing nine chiral centers was determined using a combination of NMR methods (COSY, NOESY, and ¹H-¹H NMR coupling constants) coupled with a molecular modeling study.¹⁰ The pivotal proton H-2 occurs as a doublet of doublets exhibiting strong couplings with H-1 and H-17. In agreement with the proposed relative stereochemistry, the large axial-axial coupling constant between H-1 and H-2 (J = 10.0 Hz) indicated these protons to be trans coupled, whereas the small axial-equatorial coupling constant between H-2 and H-17 (J = 6.1 Hz) required the latter protons to be cis coupled. The H-1 signal showed a proton COSY correlation only to the H₂-14 signal at $\delta_{\rm H}$ 1.73, which in turn showed no couplings with H-13. If we arbitrarily assume that H-1 is in a pseudoaxial conformation above the plane of the molecule, we can deduce that H-14 α ($\delta_{\rm H}$ 1.73) must be below the plane in a trans pseudodiaxial conformation to H-1. As H-13 showed a proton COSY correlation only with the H₂-14 signal at $\delta_{\rm H}$ 3.02 (H-14 β), which in turn showed no couplings with H-1, hence H-13 and H-14 β must lie in a nearly antiperiplanar conformation (J = 10.9 Hz), placing the acetate group at C13 above the plane in the β configuration. Strong NOESY correlations between H-2, H-14 α , and H₃-19 with H-13 established the spatial proximities of these protons on the bottom face of the molecule (Table 1). Similarly, NOESY interactions between H-7 and the H₂-9 signal at $\delta_{\rm H}$ 2.05 (H- 9β) allowed the assignment of these protons to the upper face of 1. On the other hand, pronounced NOESY correlations between H-10, H-11, and H₃-19 with the H₂-9 signal at $\delta_{\rm H}$ 2.45 (H-9 α) placed these protons below the plane,

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⁽¹⁰⁾ Lowest energy conformers were searched using MMFF force field implemented in the MacSpartan Pro program (Wavefunction, Inc.).



Figure 2. ORTEP diagram with 30% probability of ellipsoids showing the crystallographic atom-numbering scheme and solid state conformation of providencin (1). The hydrogen atoms have been omitted for clarity.

resulting in the geometry proposed. The spatial relationships of the γ -lactone, furan, epoxide, and cyclobutane rings within the macrocyclic system was further confirmed by the near absence of coupling (J < 1.0 Hz) of the $\delta_{\rm H}$ 3.99 absorption for H-11, which can be attributed to the combined electronegativity effects of vicinal trans coplanar oxygen atoms on the coupling strength of the H-10 and H-11 protons.¹¹ The dihedral angles between these protons diminish the coupling strength of each proton, reducing their mutual coupling to

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less than 1 Hz. Thus, the overall relative stereochemistry for providencin (1) is $1R^*, 2R^*, 7S^*, 8R^*, 10S^*, 11S^*, 12S^*, 13R^*, 17R^*$.

Recrystallization by slow evaporation in a mixture of 9:1 $MeOH/CHCl_3$ produced good quality crystals of **1**, which allowed a single-crystal X-ray structure analysis to be carried out, thus confirming its molecular structure. The computer-generated perspective drawing of the final X-ray model of **1**, without hydrogens, shown in Figure 2 revealed that the solution conformation is similar to that in the solid state. The absolute configuration of the molecule was not determined in the X-ray experiment. The novel carbon skeleton of providencin (**1**), which we named providenciane, represents a new class of marine-derived diterpenes. A numbering scheme that preserves the C1 to C20 numbering of the parent cembrane skeleton is proposed.

Providencin (1) displayed modest in vitro cytotoxicity against MCF7 breast cancer, NCI-H460 nonsmall cell lung cancer, and SF-268 CNS cancer. The percent of growth of the treated cells when compared to the untreated control cells was approximately 57, 39, and 94%, respectively.

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Supporting Information Available: Detailed description of experimental procedures and tables of crystal data for providencin (1) (crystal data and structure refinement, atomic coordinates, bond lengths and angles, anisotropic displacement parameters, and hydrogen coordinates). This material is available free of charge via the Internet at http://pubs.acs.org.

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